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Interactive Effect of Elevated CO₂ and Drought Stress on Leaf Anatomy in *Brassica Species*

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Author's contribution

The sole author designed, analyzed, interpreted and prepared the manuscript.

Article Information

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Original Research Article

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ABSTRACT

The aim of present study was to understand how the ultrastructure of the leaf mesophyll cells in *Brassica* leaf can be altered under elevated CO_2 by interactive effect of elevated CO_2 on leaf anatomy and ultra structure of Brassica species under moisture stress conditions. Results of the experiment revealed that the crop genotypes differ greatly in response to elevated CO_2 and moisture stress conditions. Elevated CO_2 brought about an increase in cell and chloroplast expansion in *Brassica* genotypes. Elevated CO_2 also increased the thickness of epidermis, size ofmesophyll cells, accumulation of starch and size and number of starch granules per chloroplast in *Brassica juncea* and *Brassica juncea* cultivars. These alterations in the ultra structure of cells in plants might help to plant adjustment to changing climate in the future.

Keywords: Elevated CO₂; moisture stress; chloroplast; starch granule and leaf thickness.

1. INTRODUCTION

The CO_2 concentration in the atmosphere was in a steady state at 280 mol mol⁻¹ till the preindustrial period (1850). If it continues at the present rate i.e.1.5 ppm (exponential rise of CO_2 in the atmosphere); it may be doubled by the end of 21^{st} century (Luo and Moony 1996). Atmospheric CO₂ increased to $403.3 \pm 0.1 \,\mu$ mol mol⁻¹ in 2016, approximately 145% of preindustrial level [1]. This increase will inevitably continue [2]. CO₂ has biological effect as well,

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because it serves as the main substrate for photosynthetic carbonassimilation. However, there is no consensus on the quantitative effects of increased C CO₂ on plant processes and growth, due to differences in response with stages of growth, type of species and crops and because of growth limiting environmental factors. Hence, the various researches are being carried on to study the crop response to elevated CO₂. The current increase in atmospheric CO₂ at the rate of 1.5-1.8 µmol mol⁻¹ per annum is much too small a signal for field experiment to detect a response. Therefore, various models were used for CO₂ experiments using Open Top Chambers (OTCs) and Free Air CO2 Enrichment (FACE) technologies that provide alarge input signal or pulse which allows tracing of crop responses for a short time frame. Crops responds directly to rising CO₂ through photo-synthesis and stomatal conductance [3,4], which consequently promotes crop yield [5,6]. The direct fertilization effect of rising atmospheric CO₂ is expected to offset the reduction of crop yields induced by climate change [7]. Besides the physiological and biochemical process elevated CO₂ has also effect on anatomical processes depending on species. Roger et al. [8] reported in increasing in thickness of all layers of leaf of Pinustaeda and Liquidambar styraciflu a grown in high CO₂. The difference in leaf thicknesswas reflected in difference in the ratio of leaf weight to leaf area [9]. Leadley [10], Wu et al. [11] reported that soybean plants grown in the high CO₂ treatment had thickerleaves but less palisade cell surface area per unit leaf area. Surface area of the mesophyll per unit leaf area was unaffected by CO₂. Overdieck and Ungemach [12] recorded that elevated CO₂ induced increase in density of palisade and spongy mesophyll at the expense of intercellularspaces in T. repens and Lolium perenne. Chloroplasts of CO2 enriched leaf tissues of plants were more deeply stained by crystal violate and appear eddenser. Elevated CO₂ stimulated cell production in roots cells [13]. Elevated CO₂ brought about an increase in cell and chloroplast expansion of wheat (Triticum aestivum) leaves 10 and 25%, respectively and observed that older leaves chloroplast contained larger starch deposits [14]. Similarly, Hao et al. [15], reported that elevated CO₂ increased the number and size of starch grains in chloroplasts of two cultivars of soybean. In general, simultaneous exposure to increased O3 reduced the impact of increased CO₂ [16]. The Transmission Electron Microscopy (TEM) study of leaf tissues showed a significant increase in the thickness of epidermis, size of mesophyll

cells, accumulation of starch and size and number of starch granules per chloroplast in *Brassica juncea* [17].

Changes in chloroplast density or volume which occur in plants grown in CO₂ enriched environment have been generally attributed to increased starch accumulation in the chloroplasts of tomato and soybean [18]. In view of the wide ranges of results that have been obtained through previous research further studies are in the need to understand its ramifications and connection with ongoing rise in the ambient CO₂ content particularly in timing and mechanism of plant adjustment to elevated CO₂ through anatomical modification which will significantly influence in the biomass production potential or productivity of crop Brassica species viz. Brassica juncea cv.'RH-30' and Brassica campestris cv.'Pusa Gold' under elevated CO₂ and moisture stress condition.

2. MATERIALS AND METHODS

2.1 Plant Material

Brassica cultivars *viz. Brassica juncea* cv. RH-30 and *Brassica campestris* cv. Pusa Gold were collected and grown for the present investigation.

2.2 Experimental Site and Growth Conditions

The response of both the species to elevated CO₂ was studied using Free Air CO₂ Enrichment Technology (FACE) to simulate the doubling CO₂ concentration at, IARI, New Delhi-12. The crops were grown in the field and inside the Mid Free Air CO₂ enrichment (FACE) facility in 8 m diameter circles. An elevated CO₂ concentration of 550 µmolmol⁻¹ was maintained throughout the crop growth period with the help of computerbased PID valves. There was no exogenous supply of CO₂ to the normal air under ambient field condition. Field was prepared bv recommended agronomic practices.

2.3 Cultural Practice

Farm yard manure was applied at the rate of 5 tons per hectare at the time of field preparation. The plant spacing, fertilizer application at the rate of 30+30:60:40 kg per hectare of nitrogen, phosphorus and potassium and other cultural practices were followed as reported by Uprety et al. [17].

2.4 Moisture Stress Treatment

Moisture stress treatment was given by restricting irrigation and bringing the soil moisture level between 7 and 10% compared to 22-25% under irrigated condition. All the observations were taken in triplicate for each treatment at Stage-1: vegetative (25 days after sowing), Stage-2: Flower bud initiation (45 DAS), Stage- 3: 50% flowering (60 DAS) and Stage-4: post flowering (75DAS).

2.5 Leafanatomy

For the measurement of leaf thickness, leaf segment of each treatment was initially placed in fixative FFA, comprising of formalin: acetic acid: ethyl alcohol: Water (10: 5: 50: 35V/V). It was then dehydrated in ethyl-butyl alcohol series and embedded in paraffin wax. Cross section was made on ultra-microtom and stained with safranine (0.5%. w/v) in 50 %,(v/v)alcohol. The thickness of palisade, mesophyll and combined epidermal layer were measured in terms of micrometer (μ m) [19].

2.6 Leaf Ultra structure Study

Ultra structural measurements were done on fully expanded leaf (6th node from the top) of mainstream using Transverse Electron Microscope (TEM) technology as reported by Robertson and Leech [20]. From the base of the leaf, slices of 2-3 mm thickness were cut and fixed in 2.5% glutaraldehyde (in 0.1 M phosphate buffer, pH7.2) at room temperature for 1 hr.

The leaf tissues were washed three times in 0.1 M phosphate buffer, pH 7.2, before post-fixation in 1 percent (v/v) osmium tetra oxide in 0.1 M phosphate buffer for two hours at room temperature. After three more washes in 0.1 M phosphate buffer, the tissue was dehydrated through an acetone series and embedded in Spurr' epoxy resin (TABB Laboratories Equipment Ltd., UK,) [21]. Ultra-thin sections were cut for TEM using a diamond knife. Double staining in uranyl acetate and lead citrate was done for taking observation in electron microscope.

3. RESULTS

3.1 Anatomical Characters Leaf Thickness

Elevated level of CO_2 brought about significant increase in leaf thickness (Plate 1, Table 1). The increased over the ambient plant was 39% (palisade), 34%(spongy), 36%(mesophyll), 29% (upper epidermis), 28 %(lower epidermis) and 34% (total leaf thickness). Moisture stress treatment significantly reduced the leaf thickness. The reduction was 22% (palisade), 19% (spongy), 18% (mesophyll), 16% (upper epidermis), 25% (lower epidermis) and 18 (total leaf thickness) compared to control. The stress induced reduction in Pusa gold under ambient 39% (palisade),37%(spongy),38% condition (mesophyll), 31% (upper epidermis),35% (lower epidermis), 37% (total thickness), where as reduction under elevated CO₂ condition was 24% (palisade),21% (spongy), 23% (mesophyll), 22% (upper epidermis),21% (lower epidermis), 22% (total leaf thickness). The stress induced reduction in RH-30 under ambient condition 35% (palisade), 33% (spongy), 34% (mesophyll), 25% (upper epidermis), 25% (lowerepidermis) and 35% total leaf thickness where as reduction in elevated CO₂ condition 21% (palisade), 17% (spongy), 19% (mesophyll), 17% (upper epidermis), 16% (lower epidermis) and 19% (total leaf thickness).

3.2 Leaf Cell Size Palisade Cells Size

The increased concentration of CO₂ significantly enhanced the palisade cell size in Brassica cultivars Table 2, Plate 2 The increase was 36% (length), 23% (breath) and 65% (area). The larger palisade cell was observed in leaves of RH-30 compared to Pusa gold. Moisture stress treatment significantly reduced cell size. The reduction was 38% (length) and 28% (breath) and 74% (area). The stress-induced reduction in Pusa gold under ambient condition was 35% length, 28% breath, and 53% area whereas reduction under elevated CO₂ was 22% (length), 16% (breath), and 35% (area). The stressinduced reduction in RH-30 under ambient conditionwas 28% length, 23% breath and 53% (area) where as in elevated CO₂ reduction 19% (length),12%(breath) and 30% was (area).

3.3 Spongy Cell Size

There was no any significant effect on spongy cell size in leaves of *Brassica* cultivars due to CO_2 enrichment (Table 3). The larger spongy cell was observed in leaves of Pusa gold compared toRH-30.Moisture stress treatment significantly reduced spongy cell size. The reduction was 12% (length), 12% (breath) and 21% (area). The stress-induced reduction in Pusa gold under ambient condition was 12% (length), 16% (breath) and 26% (area) however

in RH-30 was 17% (length), 17% (breath) and 19% (area).

3.4 Palisade Number

The increased concentration of CO_2 significantly enhanced (12.33%) the palisade cellnumber in *Brassica* cultivars. (Table 2).The more number of palisade cell was observed in leaves of RH-30 compared to Pusa gold. Moisture stress treatment significantly reduced (31%) number of palisade cell. The stress induced reduction in Pusa gold under ambient was 31% where it was 21% in elevated CO_2 condition. The reduction in RH-30 under ambient was 24% where as 17% in under elevated CO_2 condition.

3.5 Spongy Cell Number

There was no any effect on spongy cell number due to CO_2 enrichment (Table 3). The more number of spongy cells was observed in leaves of Pusa gold compared to RH-30. Moisture stress treatment significantly reduced (14%) number of spongy. The stress reduction in Pusa gold under ambient was 20.00% where as in RH-30 it was15%.

3.6 Chloroplast Ultra-structure

3.6.1 Mesophyll chloroplast number

The CO_2 enrichment brought about significant increase (72 %) in the chloroplast number in the mesophyll. (Table 4) The more number of chloroplast was recorded in RH-30 cultivar compared to Pusa gold. Moisture stress significantly decreased (25%) the chloroplast number in mesophyll cell. The stress induced reduction in chloroplast number under ambient was 34% whereas under elevated 22% in Pusa gold. The reduction in RH-30 under ambient was 32%, where as in elevated 19%.

3.6.2 Vascular sheath chloroplast number

The CO_2 enrichment significantly enhanced (36%) the chloroplast number in vasculars heath cell. There was no significant difference between cultivars. Moisture stress significantly decreased (25%) in the chloroplast number in vascular bundle cell. The stress induced reduction in chloroplast number under ambient was 34% whereas in elevated condition, 21% in Pusa gold.The reduction in RH-30 under ambient was 33%, where elevated CO_2 condition only 17%.

3.6.3 Number of starch granule in mesophyll chloroplast

Elevated CO₂ brought about significant increase (35%) in the number of starch granule inthe mesophyll chloroplast. Moisture stress significantly decreased (33%) the number of starch granule in the mesophyll chloroplast. The stress induced reduction in number of starch granule in chloroplast under ambient was 48% where in elevated 24% in Pusa gold. The reduction inRH-30underambientwas 46%, where elevated 21%.

3.6.4 Number of starch granule in vascular sheath chloroplast

The CO_2 enrichment significantly increased (41%) the starch granule in the vascular sheath chloroplast (Table 4). Moisture stress significantly decreased (19%) the number of starch granule in the vascular sheath chloroplast. The stress induced reduction in number of starch granule in chloroplast under ambient was 42% where as elevated condition 20% in Pusa gold. The reduction in RH-30 under ambient was 33%, where elevated condition 15%.

4. DISCUSSION

Changes in rate of cell division, cell expansion and cell cycling was reported by various scientist due to exposure to elevated CO₂ and that might alter plant structure for example an alteration in plant structure under elevated CO₂ at organ level is possibly a result of metabolic changes induced at cellular level [22]. Elevated CO₂ brought about an increasein leaf thickness, which was also more prevalent in 'RH-30', whereas stress-induced reduction in leaf thickness greatly ameliorated Plate 1. The increase of the palisade and spongy layers alongwith that of epidermal cell layers contributed to the total leaf thickness under CO_2 though increased elevated CO₂ concentration and has no significant effect on cell size and number inspongy cells but a significant increase in palisade cell size was recorded and this results were in accordance with the results of Zheng et al. [23] reported in soybean which showed spongy tissue area of 15 and 28% as CO₂ concentration increased from 400 ppm to 600 ppm. Moisturestress significantly decreased the cell size and number in spongy layer. The increase in epidermal layer was one of most important strategy of plant for

Treatments			Pusa gold						RH-30			
	Palisade (µm)	Spongy (µm)	Mesophyll (µm)	Upper epidermi s(µm)	Lower epidermis (µm)	Total (µm)	Palisade (µm)	Spongy (µm)	Mesophyll(µm)	Upper epidermis (µm)	Lower Epidermis (µm)	Total(µm)
FACEIRR	88.19	99.90	188.10	27.21	19.40	230.71	73.89	84.21	158.10	23.20	15.30	196.60
FACEMS	66.34	78.40	144.80	21.20	15.20	181.20	57.89	69.12	127.01	19.10	12.70	158.81
AMBIRR	70.23	80.40	150.60	22.200	16.10	188.90	57.94	70.14	128.08	19.20	12.80	160.08
AMBMS CDat0.05	42.21	50.21	92.40	15.10	10.40	117.90	37.21	46.90	84.11	14.30	9.6	108.01
CV.	2.11	6.21	8.89	1.86	0.62	9.14	2.11	6.21	8.89	1.86	0.62	9.14
CO2	4.32	3.14	12.58	0.70	0.31	12.98	4.32	3.14	12.58	0.70	0.31	12.98
CVxCO2	6.99	4.45	14.97	0.99	0.44	15.19	6.99	4.45	14.97	0.99	0.44	15.19
MS	2.56	5.30	6.67	1.01	0.53	7.01	2.56	5.30	6.67	1.01	0.53	7.01
CV.xMS	7.34	7.50	13.89	1.43	0.75	14.67	7.34	7.50	13.89	1.43	0.75	14.67
CO2xMS	9.77	7.49	16.76	1.53	0.85	17.45	9.77	7.49	16.76	1.53	0.85	17.45
CV.xCO2xMS	12.56	10.60	20.55	2.03	1.36	21.55	12.56	10.60	20.55	2.03	1.36	21.55

Table 1. Interactive effect of elevated CO₂ and drought stress on leaf anatomy *Brassica Species*

FACE = Free air CO₂ enrichment, IRR= Irrigated, MS= moisture stress, AMB= ambient

Table 2. Interactive effect of elevated CO2 and moisture stress in leaf palisade cell number and cellsize

Treatment		Pusa gold				RH-30		
	Number	Length(µm)	Breath(µm)	Area(µm²)	Number	Length(µm)	Breath(µm)	Area(µm²)
FACEIRR	23.00	37.40	22.60	845.24	27.00	48.50	25.90	1256.15
FACEMS	18.00	28.90	18.81	543.61	22.30	38.90	22.60	879.14
AMBIRR	19.00	29.20	19.70	575.24	21.00	39.12	22.80	891.94
AMB MS	13.00	18.90	14.10	266.49	15.90	28.0	17.40	487.20
CV.	1.01	1.67	1.93	37.96	1.01	1.67	1.93	37.96
CO2	1.58	2.22	2.45	81.35	1.58	2.22	2.45	81.35
CV.xCO2	2.56	3.78	3.47	115.04	2.56	3.78	3.47	115.04
MS	1.67	2.92	2.43	76.90	1.67	2.92	2.43	76.90
CV.xMS	2.87	4.01	3.44	108.75	2.87	4.01	3.44	108.75
CO2xMS	3.54	5.85	3.69	118.75	3.54	5.85	3.69	118.75
CV.xCO2xMS	4.09	7.44	4.86	153.80	4.09	7.44	4.86	153.80

Treatment		Pusagold					RH-30	
	Number	Length(µm)	Breath(µm)	Area(µm²)	Number	Length(µm)	Breath(µm)	Area(µm²)
FACEIRR	17.00	31.20	22.90	714.48	14.00	26.60	19.44	517.1
FACE MS	16.00	30.64	21.42	650.18	13.00	24.97	18.91	472.18
AMBIRR	18.00	32.62	24.30	792.67	15.00	28.22	22.46	533.64
AMBMS	15.00	28.62	20.30	580.98	13.00	23.42	18.44	431.86
CV.	1.5	1.56	0.62	50.44	1.5	1.56	0.62	50.44
CO ₂	NS	NS	0.87	NS	NS	NS	0.87	NS
CV.xCO ₂	0.22	NS	1.28	NS	0.22	NS	1.28	NS
MS	0.44	0.83	0.61	18.55	0.44	0.83	0.61	18.55
CV.xMS	0.63	1.13	1.02	30.87	0.63	1.13	1.02	30.87
CO ₂ xMS	0.83	2.44	1.99	42.43	0.83	2.44	1.99	42.43
CV. x CO ₂ xMS	1.21	3.10	2.84	67.54	1.21	3.10	2.84	67.54

Table 3. Interactive effect of elevated CO₂ and moisture stress leaf spongy cell number and cellsize

Table 4. Interactive effect of elevated CO₂ and drought stress ultra structure of chloroplast

Treatments	Number of chloroplastin mesophyll		Number of starch granule in mesophyll chloroplast		Number of chloroplastin Vascular sheath		Number of starch granule in vascular sheath chloroplast	
	Pusa gold	RH-30	Pusa gold	RH-30	Pusa gold	RH-30	Pusagold	RH-30
FACEIRR	7.50	8.90	3.70	3.90	3.20	3.50	3.00	3.20
FACEMS	5.80	7.20	2.80	3.10	2.50	2.90	2.40	2.70
AMBIRR	4.70	5.50	2.90	2.90	2.60	2.70	1.90	1.80
AMBMS	3.10	3.70	1.50	1.50	1.70	1.80	1.10	1.20
Cv.	84			NS		NS		NS
CO ₂	11			0.08		0.09		0.07
Var.x CO ₂	18			0.12		0.12		0.10
MS	09			0.11		0.06		0.05
Var. xMS	43			0.22		0.12		0.11
CO ₂ xMS	67			0.43		0.30		0.21
Var. x CO ₂ xMS	42			0.61		0.42		0.30

amelioration of moisture stress effect because the thicker epidermis may reduce the rate of transpiration through the accumulation of cuticular wax. These results were in conformity with that of Thomas and Harvey (1983) in *Glycin max*) and Lee *et al.* (1993) in *Betula pendula*. Uprety et al. [17] also reported at elevated CO_2 concentration has significant effect on leaf structure of *Brassica juncea* L.cv.

Line Bio-141(95) under moisture stress which revealed that CO₂ elevated to 600 µmolmol⁻¹ increased the length of epidermal cell and length of palisade parenchyma cells Ferris and Taylor (1999) in Populus demonstrated that carbon induced growth effect occurred because of the enhanced wall extensibility and the effect of wall loosening enzyme, xyloglucan endo transglycosylase (XET). Masale [24] and Zhuo et al. [13] observed increased supply of soluble carbohydrates stimulated both cell wall expansion and cell production but the mechanism remained elusive. Ferris et al. [25] reported that leaf expansion was due to the production of larger cells in their in P. nigra when they raise the atmospheric CO₂ under FACE experiment. According to Thomas and Harvey [26] differences in thickness were due to the result of increased palisade at initiation and development. According to them in soybean and sweet gum at elevated CO₂ due to higher carbon accessibility increased osmotic potential in the leaves that in turn causes expansion of cells of palisade area. Elevated CO₂ induced one more layer ofpalisade cells in the mesophyll and greater cell expansion in Brassica leaves in the present investigation.

Vaz et al. [27] reported that when leaves of Quer cussuber was exposed to elevated CO_2 it resulted in increased leaf thickness and an additional layer of palisade cells was also observed.

Uprety 2001 opined that increase in mesophyll layer served as an important approach for Brassica juncea to provide room for higher starch accumulation under high CO2. In the present study, with Brassica, the length and width of epidermal cells were increased at high CO₂ condition, indicating greater effect on cell expansion, which might be associated with an in osmotic potential increase due to accumulation of saccharides. These changes along with those in stomatal characters might help in regulating and reducing transpiration, gas exchange and also regulating osmotic

movement of cell sap to maintain turgor under moisture stress condition. Increased chloroplast size has been a reliable observation under elevated CO_2 enrichment asreported by some workers [28,17]. According to them this has been mostly attributed to increased cross-sectional width of chloroplast rather than length.

The CO₂ enrichment significantly increased the chloroplast number in both mesophyll and vascular bundle sheath of the cell in Brassica leaves but it markedly decreased under moisture stress condition. The stress induced reduction in chloroplast number significantly ameliorated in elevated CO₂ condition. Mesophyll cell had more number of chloroplast compared to the vascular bundle sheath cell and laying near the cell wall under elevated CO₂ (Plate 2). The increased chloroplast number could be related with increase in the cell size. These findings indicated that cells have got more space to develop number of chloroplast. Similarly CO₂ enrichment brought about significant increase in the starch granule number within thechloroplast both mesophyll and vascular bundle sheath cell. There was no significant difference in terms of cultivars. Moister stress decreased the starch granule number within chloroplast in both mesophyll and vascular bundle sheath cells (Plates 3 and 6).

Several workers have studied the response of higher level of atmospheric CO_2 on starchaccumulation in the leaves of various plants for example; number and size of starch increased in cucumber [29]; in *Gmelina arborea* [30] and in *Isatis indigotica* [15] grown under Free air atmospheric CO_2 enrichment facility. And same trend was also recorded [31] in the leaves of *Impatiens hawker* when treated with higher level of CO_2 in growth chamber.

The Transmission Electron Microscope (TEM) study of leaf tissues showed a significant increase in the thickness of epidermis, size of mesophyll cells, accumulation of starch and size and number of starch granules per chloroplast in *Brassica juncea* [17]. According to them the production of photo-assimilates under elevated CO_2 might have increased the sink capacity to adjust the starch granules in the chloroplast without disturbing the thylakoids membrane. The important phenomenon observed in the present study showed that CO_2 enrichment not only increased chloroplast size but also strengthened the chloroplast membrane. The elevated CO_2 increased the starch granule size and it

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deposited them to near chloroplastmembrane stretching without any damage [Plate 4.(a) & (b)]. Due to deposition of starch granule near the membrane the thylakoid and grana were protected from mechanical damages in the middle portion of chloroplast and ultimately avoid the feedback inhibition by metabolite adjustment Plate 5. The greater size and number of starch granules additionally helped the osmotic phenomenon [32,33].



Plate 1. Effect of elevated CO2 and moisture stress on leaf anatomy of *B. Campestris* cv Pusa gold and *B. juncea*, cv. RH-30 (LM100x)



Plate 2. Effect of elevated CO_2 and moisture stress on vascular bundle cell size and number of chloroplast in the leaves *Brassica campestris cv.Pusa gold Brassica juncea* cv. RH-30



Plate 3. Effect of elevated CO2 and moisture stress on chloroplast structure in the leaves of Brassica junceacv. RH-30. (EM;8200x)





FACEIIRR FACEMS A&B:Depositionofstarchgranulenearthemembranebystretc hingwithout anydamagetochloroplast machineryunderelevated CO₂



FACE IRR FACEMS A & B:Deposition of starch granule near the membrane bystretching without any damagetochloroplast machinery underelevatedCO₂





AMBIRR AMBMS C&D:Normaldepositionofstarchgranulewithinthe chloroplast

Plate 4(a). Effect of elevated CO2 and moisture stress on starch granules in the leaves of *Brassica juncea* cv.RH-30(EM 8400x.)



AMBIKK AMBMS C&D:Normaldepositionofstarchgranulewithinthe chloroplast

Plate 4(b). Effect of elevated CO2 and moisture stress on starch granule in the leaves of *Brassica campestris* cv. Pusa Gold (EM 8400x)



Plate 5. Avoidance of feedback inhibition in Brassica cultivar under elevated CO₂



FACEIRR





AMBMS

Plate 6. Effect of elevated CO₂ and moisture stress on number of mesophyll cell chloroplast in leaves of Brassica Juncea cV. RH-30 (EM; 2050x)

5. CONCLUSION

To understand the mechanism of responses to elevated CO2, an alteration in anatomical and leaf ultra-structure is very much pertinent which provides the pave for an idea about adaptation strategies of plant under future environment. Thus, in the present investigation the response of Brassica species to the interaction of elevated CO2 and moisture stress was characterized using Free Air CO₂ Enrichment technology.

Among the two cultivar cv. RH-30 showed a better adaptation by optimizing various ultrastructure such as number starch granule in chloroplast, leaf thickness, cell size etc. compared to cv. Pusa gold This data base on Brassica species might help in developing model,

Identification of cultivars and modification of cultivation and nutrient application technologies for future environments.

DISCLAIMER

The products used for this research are commonly and predominantly use products in our area of research and country. There is absolutely no conflict of interest between the authors and producers of the products because we do not intend to use these products as an avenue for any litigation but for the advancement of knowledge. Also, the research was not funded by the producing company rather it was funded by personal efforts of the authors.

COMPETING INTERESTS

Author has declared that no competing interests exist.

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