



## Essential Oil of Two Iranian Horehound Species: *Marrubium propinquum* and *Marrubium parviflorum*

Sanaz Hamedeyazdan<sup>1,2</sup>, Masoomeh Zarei<sup>3</sup>, Aida Salem<sup>3</sup>, Solmaz Asnaashari<sup>4</sup>, Fatemeh Fathiazad<sup>1\*</sup>

<sup>1</sup>Drug Applied Research Centre, Tabriz University of Medical Sciences, Tabriz, Iran.

<sup>2</sup>Department of Pharmacognosy, Faculty of Pharmacy, Tabriz University of Medical Sciences, Tabriz, Iran.

<sup>3</sup>Student's Research Committee, Faculty of Pharmacy, Tabriz University of Medical Sciences, Tabriz, Iran.

<sup>4</sup>Biotechnology Research Center, Tabriz University of Medical Sciences, Tabriz, Iran.

### Article Info

#### Article History:

Received: 7 November 2016

Accepted: 8 December 2016

ePublished: 30 June 2017

#### Keywords:

-β-caryophyllene  
-Germacrene D  
-*Marrubium parviflorum*  
-*Marrubium propinquum*  
-Oleic acid  
-α-pinene

### ABSTRACT

**Background:** Two species of genus *Marrubium* belonging to the family Lamiaceae, were studied for their volatile components.

**Methods:** The essential oils were extracted from aerial parts of the plants through hydrodistillation using a Clevenger apparatus. Later, CG and CG-MS analysis were applied to assess the chemical components of the essential oils.

**Results:** Analysis of the *M. propinquum* essential oil resulted in the identification of 22 components, representing 79.6% of the total essential oil that principally contained oleic acid (19%), β-caryophyllene (7.4%) and m-tolualdehyde (5.2%). In the case of *M. parviflorum*, 20 components were identified, representing 83% of the *M. parviflorum* essential oil, among them oleic acid (11.8%), α-pinene (10.2%) and germacrene D (9.8%) were the main compounds.

**Conclusion:** Regarding the results of this study in both essential oils after the non terpenoids, sesquiterpene hydrocarbons possessed the uppermost portion of the oils. We found some similarities and differences between *M. propinquum* and *M. parviflorum* essential oils and also in comparison with other species of genus *Marrubium* which might be due to different parameters such as agrotechnical factors.

### Introduction

Herbal medicines have been limelight in the field of drug therapy due to their fundamental beneficial applications in humans' health care. One of the effective metabolites of herbal medicines has been known as essential oils. Various parts of plants could contain essential oils, accordingly they might have been used in medicine, pharmacy and food industry.<sup>1</sup> In this regard we aimed to study essential oils of two species of genus *Marrubium* from the family Lamiaceae.

Lamiaceae with about 220 genera and nearly 4000 species mainly contains flowering plants that are frequently aromatic in all parts.<sup>2,3</sup> Genus *Marrubium* from the family lamiaceae included 49 accepted species worldwide.<sup>4</sup> In Iran, *Marrubium* (horehound) is represented with 10 species, including *Marrubium propinquum* Fisch. & C.A.Mey and *Marrubium parviflorum* Fisch. & C.A.Mey.<sup>5,6</sup> Since ancient times, *M. vulgare* (white horehound) has been accepted for remedy of several disorders such as dyspeptic complaints, loss

of appetite, cough, wound healing and as a choleric in digestive and biliary complaints.<sup>7</sup>

Phytochemical analyses of *Marrubium* spp. demonstrated that they are rich in diterpenes,<sup>8-15</sup> polyphenols, flavonoids,<sup>16</sup> steroids, saponines, phenylpropanoid esters and glycosides.<sup>1,16-22</sup>

According to previous studies, different species of genus *Marrubium* possess various therapeutic effects such as a hypoglycemic effect,<sup>23</sup> antihypertensive,<sup>24</sup> antispasmodic,<sup>25</sup> antiproliferative,<sup>26-30</sup> antioxidant,<sup>31-32</sup> hepatoprotective,<sup>33</sup> gastroprotective,<sup>34</sup> antibacterial,<sup>35-38</sup> vasorelaxant,<sup>13</sup> hypolipidemic<sup>39</sup> and antioedematogenic.<sup>40</sup>

Several studies on essential oil composition of *Marrubium* species are found in literature.<sup>41-57</sup>

Evaluating the naturally occurring volatile constituents in the essential oil of a plant would be of value in characterizing species within a genus. According to the valuable results from previous studies on *Marrubium* species, *M. propinquum* and *M. parviflorum* were selected for assessing chemical components in their essential oils.

\*Corresponding Author: Fatemeh Fathiazad, E-mail: [fathiazad@tbzmed.ac.ir](mailto:fathiazad@tbzmed.ac.ir)

©2017 The Authors. This is an open access article and applies the Creative Commons Attribution (CC BY), which permits unrestricted use, distribution, and reproduction in any medium, as long as the original authors and source are cited. No permission is required from the authors or the publishers.

## Materials and Methods

### Plant Material

Aerial parts of *Marubium propinquum* were collected from Lighvan in East Azarbaijan province, Iran, during July 2014 at the flowering stage and authenticated by Amir-Hossein Talebpour from East Azerbaijan Education and Research Center for Agriculture and Natural Resources. Likewise, aerial parts of *M. parviflorum* were collected around *Misho-Dagh* mountainous near Marand in East Azarbaijan province, Iran, during July 2015 and authenticated by Atefeh Ebrahimi from Faculty of Pharmacy, Tabriz University of Medical Sciences. The voucher specimens were kept at the Herbarium of the Faculty of Pharmacy, Tabriz University of Medical science, Iran.

### Essential oil Extraction

Air-dried plant materials of the *M. propinquum* and *M. parviflorum* were submitted to hydrodistillation using a Clevenger type apparatus to extract the essential oils for about 2 hrs. As the essential oils content was low in amount, xylene was used as an absorbing medium. The resultant essential oils were kept in dark glass bottles at 4°C until analysis by GC/MS.

### Gas Chromatography-Mass Spectrometry (GC-MS)

The essential oils were analyzed by GC-MS using a Shimadzu GC-MS-QP 5050A gas chromatograph fitted with a DB1 (methyl phenyl syloxane, 60 m x 0.25 mm i.d., 0.25 µm film thickness) capillary column. The GC was set at the following conditions with helium as the carrier gas; flow rate of 1.3 mL/min; linear velocity: 29.6 cm/s; Split ratio, 1:24; column temperature, 2 min in 50°C, 50-270 °C at 3 °C/min; injector temperature, 250 °C, and 1 µL of volume injection of the essential oil. The MS operating parameters were as follows: ionization potential, 70 eV; ion source temperature; 270 °C; quadrupole 100 °C, Solvent delay 2 min, scan speed 2000 amu/s and scan range 30-600 amu, EV voltage 3000 volts.

### Identification of the compounds

Retention indices for all compounds were determined according to the Kovats retention indices using n-alkanes series as standards. The components of the essential oils were identified by comparison of the retention indices and mass spectral data with those for the standards and by computer matching with the Wiley 229, Nist 107, Nist 21 Library, as well as by comparing the fragmentation patterns of the mass spectra with those reported in the literature. For quantification purpose, relative area percentages were obtained by FID without the use of correction factors, where the

FID detector condition was set on a duplicate of the same column applying the same operational conditions.

## Results

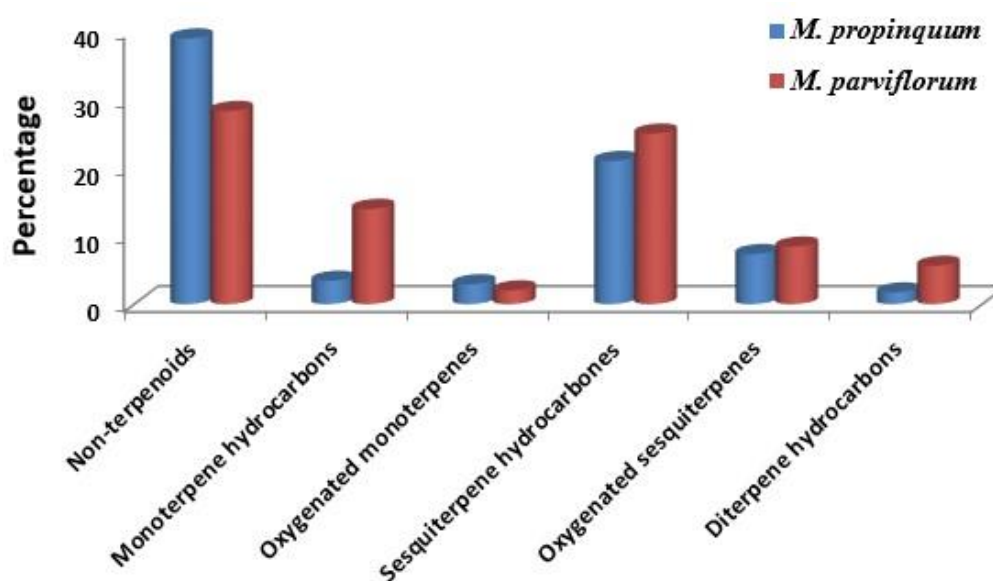
All the identified components in both essential oils are shown in Table 1, based on the order of their elution on DB1-MS column; retention indices and percentages. The GC-MS analysis of the essential oil from *M. propinquum* resulted in the identification of 22 components, representing 79.6% of the total essential oil. As shown in Figure 1, the components of *M. propinquum* essential oil were classified to non-terpenoids (38.8%), sesquiterpene hydrocarbons (20.9%), oxygenated sesquiterpenes (7.3%) monoterpene hydrocarbons (3.4 %), and oxygenated monoterpenes (2.9%).

Among the non-terpenoid compounds, m-tolualdehyde, p-tolualdehyde, and oleic acid were the dominant constituents with 5.2%, 2.9% and 19%, correspondingly. The major sesquiterpenes of *M. propinquum* essential oil were identified as β-caryophyllene (7.4%), bicyclogermacrene (4.6%), germacrene D (4.1%) and β-farnesene (3.7%). Besides, limonene and α-pinene were considered as the most frequent monoterpenes of the *M. propinquum* essential oil with the relative percentages of 1.9% and 1.4%, respectively. Considering the oxygenated monoterpenes and oxygenated sesquiterpenes, anethole (2.9%) and caryophyllene oxide (5.4%) were the most dominant components of the groupings. To sum up, essential oil of *M. propinquum* principally contained oleic acid (19%), β-caryophyllene (7.4%), m-tolualdehyde (5.2%) bicyclogermacrene (4.6%), and germacrene D (4.1%). In the case of *M. parviflorum* essential oil, 20 components were identified, representing 83% of the total essential oil that are classified to non-terpenoids (28.2%), sesquiterpene hydrocarbons (24.9%), monoterpene hydrocarbons (13.9%), oxygenated sesquiterpenes (8.4%) and oxygenated monoterpenes (2.0%). The most abundant sesquiterpenes were shown to be germacrene D (9.8%), β-caryophyllene (5.3%), β-farnesene (3%), and bicyclogermacrene (5.8%). Moreover, results showed that oleic acid (11.8%) (a monounsaturated omega-9 fatty acid) was the foremost constituent. Additionally, monoterpenes of *M. parviflorum* essential oil were rich in α-pinene, and limonene with the relative percentages of 10.2%, and 2.2%. In view of the oxygenated monoterpenes and oxygenated sesquiterpenes, verbenol (1.3%) and caryophyllene oxide (5.2%) were the leading components of the groupings. Taken as a whole, the major constituents of the *M. parviflorum* essential oil were detected to be oleic acid (11.8%), followed by α-pinene (10.2%), germacrene D (9.8%), bicyclogermacrene (5.8%) and β-caryophyllene (5.3%).

**Table 1.** Chemical constituent of the essential oils obtained from aerial parts of *M. propinquum* and *M. parviflorum*.

No.	Compounds	RI <sup>a</sup>	<i>M. propinquum</i> (%)	<i>M. parviflorum</i> (%)	Identification Method
1	$\alpha$ -Pinene	917	1.4	<b>10.2</b>	GC/MS, RI
2	1-Octen-3-ol	-	1.7	5.1	GC/MS
3	Sabinene	933	-	1.5	GC/MS, RI
4	Limonene	960	1.9	2.2	GC/MS, RI
5	m-Tolualdehyde	-	5.2	2.7	GC/MS
6	p-Tolualdehyde	-	2.9	1.6	GC/MS
7	Linalool	992	-	0.7	GC/MS, RI
8	Verbenol	1019	-	1.3	GC/MS, RI
9	Anethole	1078	2.9	-	GC/MS, RI
10	$\beta$ -Bourbenene	1392	-	1.0	GC/MS, RI
11	$\beta$ -Caryophyllene	1410	<b>7.4</b>	5.3	GC/MS, RI
12	$\beta$ -Farnesene	1424	3.7	3.0	GC/MS, RI
13	Germacrene D	1440	4.1	<b>9.8</b>	GC/MS, RI
14	Bicyclogermacrene	1448	4.6	5.8	GC/MS, RI
15	Beta-bisabolene	1452	1.0	-	GC/MS, RI
16	Spathulenol	1484	1.9	2.5	GC/MS, RI
17	Caryophyllene oxide	1488	<b>5.4</b>	5.2	GC/MS, RI
18	Eicosane	-	1.7	-	GC/MS
19	$\alpha$ - Bisabolol	1532	-	0.7	GC/MS, RI
20	Oleic acid	1821	<b>19</b>	<b>11.8</b>	GC/MS, RI
21	Phytol	-	4.6	5.6	GC/MS
22	Linoleic acid	1897	1.7	-	GC/MS, RI
23	Heneicosane	-	5.2	-	GC/MS
24	Tricosane	-	1.9	4.3	GC/MS
25	Tetracosane	-	1.2	2.7	GC/MS
<b>Total identified</b>			<b>79.6</b>	<b>83</b>	
<b>Non-terpenoids</b>			<b>38.8</b>	<b>28.2</b>	
<b>Monoterpene hydrocarbons</b>			<b>3.4</b>	<b>13.9</b>	
<b>Oxygenated monoterpenes</b>			<b>2.9</b>	<b>2.0</b>	
<b>Sesquiterpene hydrocarbones</b>			<b>20.9</b>	<b>24.9</b>	
<b>Oxygenated sesquiterpenes</b>			<b>7.3</b>	<b>8.4</b>	
<b>Oxygenated diterpenes</b>			<b>1.7</b>	<b>5.6</b>	
<b>Unidentified</b>			<b>20.4</b>	<b>17</b>	

RI<sup>a</sup> is the Retention Index relative to C8–C20 n-alkanes on the DB-1 column.



**Figure 1.** Comparison between the percentage of identified chemical groups of *M. propinquum* and *M. parviflorum* essential oils.

## Discussion

According to the data in table 1, both *M. propinquum* and *M. parviflorum* were assigned as higher content of non-terpenoid compounds plants. Previous reports on the essential oils indicated that a higher numbers of sesquiterpenes were present in different species of *Marrubium* genus.

A review on prior studies on *M. parviflorum* essential oil from different localities represented various compounds as the foremost constituents of the essential oils. For instance, bicyclogermacrene, germacrene D and  $\beta$ -caryophyllene with virtual percentages of 26.3%, 21.5% and 15.6% were specified in *M. parviflorum* essential oil collected from Khalkhal, Iran. In another report on the essential oil of *M. parviflorum* from Nevsehir Turkey, hexadecanoic acid (15.4%), germacrene D (11.1%) and  $\beta$ -caryophyllene (10.0%) were determined as the major chemical compounds.<sup>54,55</sup> Whereas in the present study, oleic acid (11.8%),  $\alpha$ -pinene (10.2%) and germacrene D (9.8%) were specified as the main compounds of the *M. parviflorum* essential oil which was collected from Marand region, Iran.

Based on the findings of our study, the major constituents for *M. propinquum* was revealed to be  $\beta$ -caryophyllene (7.4%), bicyclogermacrene (4.6%) and germacrene D (4.1%) which was different from the previous report in the literature. In the study conducted by Tajbakhsh *et al* on *M. propinquum* essential oil, it was reported that the plant collected from the suburb of Gadouk Mazandaran province in northern Iran contained mainly  $\beta$ -farnesene (43.8%),  $\beta$ -caryophyllene (20.1%) and germacrene D (4.115.8%).<sup>56</sup>

Having compared the essential oils of *M. propinquum* and *M. parviflorum*, we could bring about the prime difference between *M. propinquum* and *M. parviflorum* essential oil due to the monoterpenoid groupings, as regards in the Figure 1. *M. parviflorum* essential oil was rich in monoterpene hydrocarbons (13.9%) and  $\alpha$ -pinene with the relative percentages of 10.2% was determined as a chief monoterpene. On the contrary, in *M.*

*propinquum*, monoterpene hydrocarbons were relatively in a lower amount of 3.4% of the essential oil. In comparison, in essential oil of *M. propinquum* not only allocated  $\alpha$ -pinene and limonene in higher percentages but also contained a monoterpene hydrocarbons, sabinene, which was not identified in *M. propinquum*. In addition, the constituents of the oxygenated monoterpenes were completely different in the relative essential oils. So that, linalool and verbenol were present as oxygenated monoterpenes in *M.*

*parviflorum* whereas, anethole was the main component in *M. propinquum* essential oil. Nonetheless, in both essential oils after the non

terpenoids, sesquiterpene hydrocarbons possessed the uppermost portion of the oils. On the whole, we could find much similar compounds in different quantities accounting for 79.6% of *M. propinquum* essential oil and 83% of *M. parviflorum* essential oil.

Upon earlier reports germacrene D was ascertained as the major sesquiterpene hydrocarbone in *M. anisodon* (44.2%) and *M. incanum* Desr. (28.75-32.14%).<sup>52,56</sup> Likewise, bicyclogermacrene, one of the major compounds of the sesquiterpene hydrocarbons in *M. propinquum* and *M. parviflorum* essential oils, was usually absent in the essential oil of *M. vulgare*, the most famous species of *Marrubium* genus. As we know, geographical, seasonal and climatic issues would commonly influence the chemical composition of the essential oils within various populations of these species.

## Conclusion

Concerning the results of this study we could find some similarities and differences between *M. propinquum* and *M. parviflorum* essential oils and also in comparison with other species of *Marrubium* genus. Generally, mentioned similarities and differences might be attributed to both intrinsic parameters of the plants (genetic, growth stage, etc.) and extrinsic factors such as climatic conditions, seasonal variations, environmental issues, distillation processes and etc.

## Acknowledgments

The authors would like to thank Drug Applied Research Centre, Tabriz University of Medical Sciences, Iran for financial support of this study.

## Conflict of interests

The authors claim that there is no conflict of interest.

## References

1. Meyre-Silva C, Cechinel-Filho V. A review of the chemical and pharmacological aspects of the genus *marrubium*. *Curr Pharm Des*. 2010;16(31):3503-18. doi:10.2174/138161210793563392
2. Hadley SK, Petry JJ. Medicinal herbs: A primer for primary care. *Hosp Pract*. 1999;34(6):105-23. doi:10.3810/hp.1999.06.151
3. Naghibi F, Mosaddegh M, Mohammadi Motamed M, Ghorbani A. Labiatae family in folk medicine in iran: From ethnobotany to pharmacology. *Iran J Pharm Res*. 2005;4(2):63-79.
4. Lamiaceae. The Plant List. <http://www.theplantlist.org/1.1/browse/A/Lamiaceae>. Accessed 1st January 2013.
5. Rechinger KH, Hedge IC. *Flora Iranica*. Graz: Akademische Druck and Verlagsanstalt; 1982.



- p. 439.
6. Mozaffarian V. A dictionary of iranian plant names. Tehran: Farhang Moaser; 2003. p. 338.
  7. Gruenwald J, Brendler T, Jaenicke C. PDR for herbal medicines. Montvale, N.J.: Medical Economics Company; 2000. p. 401
  8. Zhang J-S, Zou Y-H, Zhao J-J, Chen Y, Bao J-M, Tang G-H. Three new diterpenoids from *marrubium aschersonii*. *Phytochem Lett.* 2016;16:241-4. doi:10.1016/j.phytol.2016.05.004
  9. Hammami S, Li Z, Huang M, El Mokni R, Dhaouadi H, Yin S. New bioactive labdane diterpenoids from *marrubium aschersonii*. *Nat Prod Res.* 2016;30(19):2142-8. doi:10.1080/14786419.2016.1143828
  10. Argyropoulou C, Karioti A, Skaltsa H. Labdane diterpenes from *marrubium thessalum*. *Phytochemistry.* 2009;70(5):635-40. doi:10.1016/j.phytochem.2009.03.011
  11. Karioti A, Heilmann J, Skaltsa H. Labdane diterpenes from *marrubium velutinum* and *marrubium cylleneum*. *Phytochemistry.* 2005;66(9):1060-6. doi:10.1016/j.phytochem.2005.02.029
  12. El Bardai S, Wibo M, Hamaide MC, Lyoussi B, Quetin-Leclercq J, Morel N. Characterisation of marrubenol, a diterpene extracted from *marrubium vulgare*, as an l-type calcium channel blocker. *Br J Pharmacol.* 2003;140(7):1211-6. doi:10.1038/sj.bjp.0705561
  13. El Bardai S, Morel N, Wibo M, Fabre N, Llabres G, Lyoussi B, et al. The vasorelaxant activity of marrubenol and marrubiin from *marrubium vulgare*. *Planta Med.* 2003;69(1):75-7. doi:10.1055/s-2003-37042
  14. Çitoğlu GS, Aksit F. Occurrence of marrubiin and ladanein in *marrubium trachyticum* boiss. From turkey. *Biochem Syst Ecol.* 2002;30(9):885-6. doi:10.1016/s0305-1978(01)00148-x
  15. Yousefi K, Hamedeyazdan S, Torbati M, Fathiazad F. Chromatographic fingerprint analysis of marrubiin in *marrubium vulgare* l. Via hptlc technique. *Adv Pharm Bull.* 2016;6(1):131-6. doi:10.15171/apb.2016.019
  16. Karioti A, Skaltsa H, Heilmann J, Sticher O. Acylated flavonoid and phenylethanoid glycosides from *marrubium velutinum*. *Phytochemistry.* 2003;64(2):655-60. doi:10.1016/s0031-9422(03)00242-5
  17. Çaliş İ, Hosny M, Khalifa T, Rüedi P. Phenylpropanoid glycosides from *marrubium alysson*. *Phytochemistry.* 1992;31(10):3624-6. doi:10.1016/0031-9422(92)83740-p
  18. Rigano D, Formisano C, Basile A, Lavitola A, Senatore F, Rosselli S, et al. Antibacterial activity of flavonoids and phenylpropanoids from *marrubium globosum* ssp. *Libanoticum*. *Phytother Res.* 2007;21(4):395-7. doi:10.1002/ptr.2061
  19. Agbo MO, Uzor PF, Nneji UNA, Odurukwe CUE, Ogbatue UB, Mbaaji EC. Antioxidant, total phenolic and flavonoid content of selected nigerian medicinal plants. *Dhaka Univ J Pharm Sci.* 2015;14(1):35-41. doi:10.3329/dujps.v14i1.23733
  20. Argyropoulou A, Samara P, Tsitsilonis O, Skaltsa H. Polar constituents of *marrubium thessalum* boiss. & heldr.(lamiaceae) and their cytotoxic/cytostatic activity. *Phytother Res.* 2012;26(12):1800-6. doi:10.1002/ptr.4654
  21. Kurbatova N, Muzychkina R, Mukhitdinov N, Parshina G. Comparative phytochemical investigation of the composition and content of biologically active substances in *marrubium vulgare* and *M. alternidens*. *Chem Nat Compd.* 2003;39(5):501-2. doi:10.1023/b:conc.0000011128.64886.f4
  22. Sahpaz S, Garbacki N, Tits M, Bailleul F. Isolation and pharmacological activity of phenylpropanoid esters from *marrubium vulgare*. *J Ethnopharmacol.* 2002;79(3):389-92. doi:10.1016/s0378-8741(01)00415-9
  23. Boudjelal A, Henchiri C, Siracusa L, Sari M, Ruberto G. Compositional analysis and in vivo anti-diabetic activity of wild algerian *marrubium vulgare* L. Infusion. *Fitoterapia.* 2012;83(2):286-92. doi:10.1016/j.fitote.2011.11.005
  24. Anwar MA, Al Disi SS, Eid AH. Anti-hypertensive herbs and their mechanisms of action: Part ii. *Front Pharmacol.* 2016;7:1-25. doi:10.3389/fphar.2016.00050
  25. Rigano D, Aviello G, Bruno M, Formisano C, Rosselli S, Capasso R, et al. Antispasmodic effects and structure-activity relationships of labdane diterpenoids from *marrubium globosum* ssp. *Libanoticum*. *J Nat Prod.* 2009;72(8):1477-81. doi:10.1021/np9002756
  26. Hamedeyazdan S, Fathiazad F, Sharifi S, Nazemiyeh H. Antiproliferative activity of *marrubium persicum* extract in the mcf-7 human breast cancer cell line. *Asian Pac J Cancer Prev.* 2012;13(11):5843-8. doi:10.7314/apjcp.2012.13.11.5843
  27. Yamaguchi K, Liggett JL, Kim N-C, Baek SJ. Anti-proliferative effect of horehound leaf and wild cherry bark extracts on human colorectal cancer cells. *Oncol Rep.* 2006;15(1):275-81. doi:10.3892/or.15.1.275
  28. Karioti A, Skopeliti M, Tsitsilonis O, Heilmann J, Skaltsa H. Cytotoxicity and immunomodulating characteristics of labdane diterpenes from *marrubium cylleneum* and *marrubium velutinum*. *Phytochemistry.*

- 2007;68(11):1587-94.  
doi:10.1016/j.phytochem.2007.03.027
29. Hamedeyazdan S, Sharifi S, Nazemiyeh H, Fathiazad F. Evaluating antiproliferative and antioxidant activity of *marrubium crassidens*. *Adv Pharm Bull.* 2014;4(5):459-64. doi:10.5681/apb.2014.068
30. Marrelli M, Conforti F, Formisano C, Rigano D, Senatore F, Bruno M, et al. Labdane diterpenoids from *marrubium globosum* ssp. *Libanoticum* as inhibitors of proliferation of melanoma cells. *Congresso Interdisciplinare sulle Piante Medicinali*; 2012.
31. Weel KG, Venskutonis PR, Pukalskas A, Gruzdienė D, Linssen JP. Antioxidant activity of horehound (*marrubium vulgare* L) grown in lithuania. *Eur J Lipid Sci Technol.* 1999;101(10):395-400. doi:10.1002/(sici)1521-4133(199910)101:10<395::aid-lipi395>3.0.co;2-1
32. Matkowski A, Piotrowska M. Antioxidant and free radical scavenging activities of some medicinal plants from the lamiaceae. *Fitoterapia.* 2006;77(5):346-53. doi:10.1016/j.fitote.2006.04.004
33. Ettaya A, Dhibi S, Samout N, Elfeki A, Hfaiedh N. Hepatoprotective activity of white horehound (*marrubium vulgare*) extract against cyclophosphamide toxicity in male rats. *Can J Physiol Pharmacol.* 2016;94(4):441-7. doi:10.1139/cjpp-2015-0405
34. Paula de Oliveira A, Santin JR, Lemos M, Klein Júnior LC, Couto AG, Meyre da Silva Bittencourt C, et al. Gastroprotective activity of methanol extract and marrubiin obtained from leaves of *marrubium vulgare* L.(lamiaceae). *J Pharm Pharmacol.* 2011;63(9):1230-7. doi:10.1111/j.2042-7158.2011.01321.x
35. Ulukanlı Z, Akkaya A. Antibacterial activities of *marrubium catarifolium* and *phlomis pungens* var. *Hirta* grown wild in eastern anatolia, turkey. *Int J Agric Biol.* 2011;13(1):105-9.
36. Masoodi MH, Ahmed B, Zargar IM, Khan SA, Khan S, Singh P. Antibacterial activity of whole plant extract of *marrubium vulgare*. *Afr J Biotechnol.* 2008;7(2):86-7.
37. Molina-Salinas GM, Ramos-Guerra MC, Vargas-Villarreal J, Mata-Cárdenas BD, Becerril-Montes P, Said-Fernández S. Bactericidal activity of organic extracts from *flourensia cernua* dc against strains of *mycobacterium tuberculosis*. *Arch Med Res.* 2006;37(1):45-9. doi:10.1016/j.arcmed.2005.04.010
38. Petrović S, Pavlović M, Maksimović Z, Milenković M, Couladis M, Tzakouc O, et al. Composition and antimicrobial activity of *marrubium incanum* desr.(lamiaceae) essential oil. *Nat Prod Commun.* 2009;4(3):431-4.
39. Ibrahim AY, Hendawy SF, Elsayed AA, Omer EA. Evaluation of hypolipidemic *marrubium vulgare* effect in triton wr-1339-induced hyperlipidemia in mice. *Asian Pac J Trop Med.* 2016;9(5):453-9. doi:10.1016/j.apjtm.2016.03.038
40. Stulzer HK, Tagliari MP, Zampirolo JA, Cechinel-Filho V, Schlemper V. Antioedematogenic effect of marrubiin obtained from *marrubium vulgare*. *J Ethnopharmacol.* 2006;108(3):379-84. doi:10.1016/j.jep.2006.05.023
41. Golmakani H, Rabbani Nasab H, Sharifan M, Kamali H, Yadollahi A. The essential oil composition and antibacterial activity of *marrubium duabense* murata from north khorassan province, iran. *J Essent Oil Bear Plants.* 2016;19(4):963-71. doi:10.1080/0972060x.2014.998722
42. Golparvar AR, Hadipanah A, Mehrabi AM, Armin A. Chemical composition of the essential oil from the leaves of *marrubium vulgare* L. From iran. *J Herbal Drugs.* 2015;6(1):1-5.
43. Grassia A, Senatore F, Arnold NA, Russo A, Piozzi F, Rigano D, et al. Chemical composition and antimicrobial activity of the essential oils from aerial parts of two *marrubium* sp.(lamiaceae) growing wild in lebanon. *Pol J Chem.* 2006;80(4):623-8.
44. Hamdaoui B, Wannas WA, Marrakchi M, Brahim NB, Marzouk B. Essential oil composition of two tunisian horehound species: *Marrubium vulgare* L. And *marrubium aschersonii* magnus. *Journal of Essential Oil Bearing Plants.* 2013;16(5):608-12. doi:10.1080/0972060x.2013.854492
45. Hamedeyazdan S, Asnaashari S, Fathiazad F. Chemical composition of the essential oil from *marrubium persicum* ca mey.(lamiaceae). *Pharm Sci.* 2013;19(2):35-8.
46. Kadri A, Zarai Z, Békir A, Gharsallah N, Damak M, Gdoura R. Chemical composition and antioxidant activity of *marrubium vulgare* L. Essential oil from tunisia. *Afr J Biotechnol.* 2011;10(19):3908-14. doi:10.5897/AJB11.301
47. Laouer H, Yabrir B, Djeridane A, Yousfi M, Beldovini N, Lamamra M. Composition, antioxidant and antimicrobial activities of the essential oil of *marrubium deserti*. *Nat Prod Commun.* 2009;4(8):1133-8.
48. Kırimer N, Kürkçüoğlu M., Akgül G, Başer KHC, Mahmoud AA. Composition of the essential oil of *marrubium anisodon* c. Koch of turkish origin. *Rec Nat Prod* 2015;9(2):234-6.
49. Said-Al Ahl HA, Gendy AS, Mahmoud AA, Mohamed HF. Essential oil composition of

- marrubium vulgare* L. Cultivated in egypt. Int J Plant Res. 2015;1(4):138-41.
50. Saleh M, Glombitza K. Volatile oil of *marrubium vulgare* and its anti-schistosomal activity. Planta Med. 1989;55(1):105. doi:10.1055/s-2006-961873
51. Saleh M, Sarg T, Metwally A, Rakha A. Chemical constituents from *marrubium alysson*. Planta Med. 1981;41(2):202-3. doi:10.1055/s-2007-971702
52. Zawislak G. Comparison of chemical composition of the essential oil from *marrubium vulgare* L. And *m. Incanum* Desr. During the second year of cultivation. Acta Agrobot. 2015;68(1):59-62. doi:10.5586/aa.2015.002
53. Zawislak G. The chemical composition of the essential oil of *marrubium vulgare* L. From Poland. Farmacia. 2012;60(2):287-92.
54. Bal Y, Kaban S, Kirimer N, Baser KH. Composition of the essential oil of *marrubium parviflorum* Fisch. et Mey. Subsp. Oligodon (Boiss.) Seybold. J Essent Oil Res. 1999;11(3):300-2. doi:10.1080/10412905.1999.9701138
55. Khanavi M, Ghasemian L, Hosseiny Motlagh E, Hadjiakhoondi A, Shafiee A. Chemical composition of the essential oils of *marrubium parviflorum* Fisch. & Ca Mey. and *marrubium vulgare* L. From Iran. Flavour Fragr J. 2005;20(3):324-6. doi:10.1002/ffj.1425
56. Tajbakhsh M, Khalilzadeh MA, Rineh A, Balou J. Essential oils of *marrubium anisodon* C. Koch and *marrubium propinquum* Fisch. Et C A Mey., growing wild in Iran. J Essent Oil Res. 2008;20(2):161-2. doi:10.1080/10412905.2008.9699980
57. Hamedeyazdan S, Asnaashari S, Fathiazad F. Characterization of non-terpenoids in *marrubium crassidens* Boiss. Essential oil. Adv Pharm Bull. 2013;3(2):429-32. doi:10.5681/apb.2013.069