



Amazon and Brazilian Polyherbal Preparation for Treatment of Necrotizing Fasciitis

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Authors' contributions

This work was carried out in collaboration between both authors. Both authors read and approved the final manuscript.

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Case Study

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ABSTRACT

Necrotizing fasciitis (flesh-eating disease) is a rare but life-threatening condition. We report one case of severe necrotizing fasciitis affecting the leg at risk of being amputated due to infection, before and after treatment with medicinal plants which have empirical broad-spectrum antibiotic and anti-inflammatory properties. All the medical attempts with conventional antibiotics failed and we had to act rapidly in order to cure the infection and prevent the amputation. These results suggest that certain medicinal plants could represent a treatment option and that further studies are considered.

Keywords: Phytotherapy; wound; necrotizing fasciitis; infectious disease; antibiotic resistance; diabetic foot.

1. INTRODUCTION

Necrotizing Fasciitis (NF) is an uncommon complicated infection (3 cases per 100,000 people-year) of the soft tissues, often associated with life-threatening conditions due to severe

sepsis, if early medical treatment is not performed [1,2].

Medicinal plants therapeutical potential has been considered by mankind since ancient Egypt. Sometimes, these natural medications serve as

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therapeutic alternatives or, in some cases, as the only effective treatment. A combination of nine plants: *Psidium guajava* (guava tree), *Nicotiana tabacum* (tobacco), *Anacardium occidentale* L. (cashew tree), *Caesalpinia ferrea* Martius (Brazilian Ironwood), *Schinus terebinthifolius* Raddi (aroeira or Brazilian pepper tree), *Hymenaea courbaril* L. (jatobá), *Rubus sellowii*. (black capinuriba or Brazilian Rubus), *Ischnosiphon polyphyllus* (arumã), *Jatropha gossypifolia* (Pinhão-Roxo or Pião-Roxo) was used to design a protocol in order to regress and cure necrotizing fasciitis. This decoction has shown antibiotic and anti-inflammatory properties with no side effects based on the scientific literature that reports experiments carried out *in vitro* and/or *in vivo* with these plants [3-15].

2. CASE REPORT

A 61-year-old white female non-diabetic patient with necrotizing fasciitis on her leg. On November 16th, 2017, she consulted a physician who prescribed her ciprofloxacin 500mg (1 tablet – twice a day or b.i.d. = "bis in die" in Latin), dipyrone or metamizole 1 gram (1 tablet - 4 times a day or q.i.d. = "quater in die" in Latin), and clindamycin 300mg (1 tablet - 4 times a day or q.i.d.).

The treatment was prolonged during 60 days (until January 15th 2017) without any positive results (after 2 months the wound was worse). The patient complained of pain in her belly. Antibiotic causes changes in intestinal flora,

which justifies such pain. Then, the physician suggested her the debridement technique using fly larvae (*Cryomya megacephala*). Neither treatment produced regression of the necrotizing fasciitis. The patient noticed that the lesion was worse (Fig. 2). As final solution another physician told her that the leg would be amputated.

The patient stopped taking antibiotics. Desperate, she came to our laboratory with her medical report. Immediately, we prescribed her probiotics, during 1 week, in order to restore the intestinal flora. After we examined her leg, we prescribed the following:

Solution 1 (topical use only) 7:00 AM, 10:00 AM, 1PM, 3PM, 5PM, 7PM, 9PM: *Jatropha gossypifolia* (popular names: Pião-Roxo or Pinhão-Roxo). Boil 50 grams in water (1000 ml) for 5 to 10 minutes. Percolate this solution. Wash the wound with this solution. Put leaves (clean and sanitized) on the wound. Place gauze over the leaves for 2 hours. After 2 hours remove the gauze.

Solution 2 (topical use only) 9:00 AM: A combination of *Nicotiana tabacum* (tobacco) leaves (50 grams), naphthalene (mothballs) (100 grams), *Libidibia ferrea*, formerly *Caesalpinia ferrea* (popular name: Brazilian Ironwood) leaves (50 grams) in water (1,000 ml). This mixture was boiled during 10 minutes and then percolated. After cooled, this decoction was used to wash the wound in the morning.



Fig. 1. Necrotizing fasciitis on left leg before starting the treatment with conventional antibiotics

Note: Tobacco leaves cause burning sensation and naphthalene have the function of softening this sensation.

Solution 3 (topical use only) 1:00 PM: *Anacardium occidentale* (cashew tree bark) (50 grams) + *Psidium guajava L.* (guava tree bark) (50 grams) in water (1,000 ml). Boil the water and percolate the solution (decoction). Use this "tea" to wash the wound.

Solution 4 (topical use only) 5:00 PM: *Schinus terebinthifolius* (common names: Brazilian peppertree, aroeira, rose pepper) (50 grams) and *Hymenaea courbaril* (common names: jatobá, courbaril, Stinkingtoe, and West Indian locust) (50 grams) in water (1,000 ml). Boil the water and percolate the solution (decoction). Use this "tea" to wash the wound.

Solution 5 (topical use only) 9:00 PM: *Rubus sellowii* (black capinuriba) (50 grams) and arumã

bark (50 grams) in water (1,000 ml). Boil the water and percolate the solution (decoction). Use this "tea" to wash the wound.

Repeat this procedure between washes with solutions from other plants.

Solution 6 (oral use only) 1:00 PM: Ten guava tree leaves (well washed) in 250 ml of water. After boiling ten minutes, wait for it to cool down and it is ready to be drunk.

The intense pain, disproportionate to the phlogistic signs present, and the rapid recovery of bullous lesions, with an area of necrosis of deeper tissues and fascia, reinforce the hypothesis of necrotizing fasciitis (3).



Fig. 2. After 60 days of antibiotic treatment (clindamycin and ciprofloxacin) and debridement the lesion got worse



A



B

Fig. 3. A) Skin lesion after 15 days of phytotherapy treatment (left photo) on February 1st 2015; B) 28 days of treatment (February 13th 2015 - lesion completely cured) (right photo)

After three days of treatment the intense pain subsided. After 15 days of treatment, an improvement was noticed (Fig. 3A) and 23 days of treatment (lesion was completely cured).

3. DISCUSSION

Natural products from Brazilian and Amazon plants have been the resource of new compounds with antimicrobial effects.

As most cases of necrotizing fasciitis are polymicrobial, empirical broad-spectrum antibiotic coverage should be administered [4].

As the medical report of the patient was not describing the type of necrotizing fasciitis, we had to prescribe plants that produce compounds with antibiotic effect against the microorganisms which cause the main types (I, II and III): *Clostridium perfringens*, *Staphylococcus aureus*, *Escherichia coli*, haemolytic group A streptococcus, staphylococci including methicillin-resistant strains/MRSA.

Due to the urgency of the case, our strategy was to combine 9 plants that have a broad bactericidal, fungicidal and anti-inflammatory effect. Two of them (Guava and cashew) in addition to the antibiotic effect against various bacteria, have effect against *Clostridium ssp.* Since two broad-spectrum antibiotics (clindamycin and ciprofloxacin) were tested for 60 days and no success has been achieved, we assume that we could be facing a necrotizing fasciitis type III (which is caused by *Clostridium ssp.*)

Due to high risk of amputation, the “war strategy” was to prescribe as many plants as possible in order to obtain the widest antimicrobial effect; so the wound could be healed in a short period of time. The explanation for prescribing such plants is the following:

Jatropha gossypifolia (Family: Euphorbiaceae): Although is a native plant from Brazil, it has also been cultivated in many parts of Singapore, Burma, and India, and used in many countries as a popular medicine for various diseases. Jatrophenone (macrocyclic bioactive diterpene) is an isolated compound from this plant with significant antimicrobial activity against *Staphylococcus aureus* and its activity was comparable to that of the standard compound, penicillin G [5].

Nicotiana tabacum: The plant, now raised for commercial tobacco production, is from South America. Researchers have been reported that antibacterial property could be due to pyridine, 3-(1-methyl-2-pyrrolidinyl) - (S) with a broad spectrum of activity against several microorganisms including *Staphylococcus aureus* (6, 7) *Candida albicans* and *Streptococcus pyogenes* [8].

Libidibia ferrea (formerly Caesalpinia ferrea): In the Amazon region of Brazil; the fruit of this plant is widely used as an antimicrobial and healing medicine for infections [9]. Some compounds are known to be responsible for biological activity, such as saponins and phenolic [10], catechins which are active against Gram-positive bacteria [11].

Anacardium occidentale: Cashew is a multipurpose tree of the Amazon which produces many resources and products. The bark and leaves of the tree are used medicinally and it has antibacterial activity against *Escherichia coli*, *Pseudomonas aeruginosa* [12] and *Clostridium perfringens* [13].

Psidium guajava: The guava tree is a small tree native to Central and South America [14]. This plant contains catechin and it is active against Gram-positive bacteria [11]. Garcia and collaborators [15] describe that the methanol extract and the aqueous extract of guava leaves are effective inhibitors of the production of spores and enterotoxins of *Clostridium perfringens* type A. Beyond this, this plant has antimicrobial effect against *Staphylococcus sp.*, *Shigella sp.*, *Salmonella sp.*, *Escherichia coli*, *Pseudomonas sp.*, different types of fungi, yeast (*Candida*), amoebas and *Plasmodium* [16]. Guava tree leaves tea prepared by the infusion method were successfully tested against clinical influenza A (H1N1) [17]. In addition, there is a Chinese patent on the potential of guava tree leaves to be effective against coronavirus [18, 19] and anticoagulant effect [20,21].

Schinus terebinthifolius Raddi: Popularly known as “Brazilian pepper”, is an evergreen, pioneer and indigenous plant from Brazil [22]. A lectin was isolated from leaves and it has shown to be active against *Staphylococcus aureus*, *Escherichia coli*, *Klebsiella pneumoniae*, *Proteus mirabilis*, *Pseudomonas aeruginosa* and *Salmonella enteritidis* [23]. Essential oil of ripened fruits has toxicity against the growth of *Acinetobacter baumannii*, *Bacillus subtilis*,

Escherichia coli, *Micrococcus flavus*, *Pseudomonas aeruginosa*, *Sarcina lutea*, *Staphylococcus aureus* [24], *Klebsiella oxytoca*, *Corynebacterium* sp., *Enterobacter* sp., *Enterobacter agglomerans*, *Nocardia* sp. and *Streptococcus* group D [25].

Hymenaea courbaril L. is a tree common in South America Caribbean and Central America that is used for furniture, flooring, and decoration. It was extract by Da-Costa and colleagues (2014) a compound called fisetin which has therapeutic potential against fungal infections with low toxicity for animal cells [26].

Rubus sellowii (black capinuriba), is small fruit that grows in a temperate climate, presents an attractive color that varies from red to blue, due to the high anthocyanin and carotenoid content which are the major natural pigments found in such fruits. The major carotenoids found in blackberry were all-trans- β -carotene and all-trans-lutein, making up 39.6% and 28.2% of the total, respectively [10,27].

Fig. 1 shows necrotizing fasciitis before starting the treatment with conventional antibiotics. Fig. 2 shows that the lesion got worse after 60 days using clindamycin and ciprofloxacin.

4. CONCLUSION

Natural products derived from Brazilian and Amazon plants have provided a source of new antimicrobial compounds. The findings point to the possibility of some medicinal plants as a therapeutic option and call for further research.

CONSENT

As per international standard or university standard, patients' written consent has been collected and preserved by the author(s).

ETHICAL APPROVAL

It is not applicable.

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

1. Costa IMC, Cabral ALSV, de Pontes SS, de Amorim JF. Necrotizing fasciitis: new

insights with a focus on dermatological aspects. *An Bras Dermatol*. 2004;79:211-224.

2. Misiakos EP, Bagias G, Patapis P, Sotiropoulos D, Kanavidis P, Machairas A. Current concepts in the management of necrotizing fasciitis. *Front Surg*. 2014;1:36.
3. Biswas B, Rogers K, McLaughlin F, Daniels D, Yadav A. Antimicrobial Activities of Leaf Extracts of Guava (*Psidium guajava* L.) on Two Gram-Negative and Gram-Positive Bacteria. *Int J Microbiol*. 2013;2013:746165.
4. Hasham S. Necrotising fasciitis. *BMJ*. 2005;330:830–833.
5. Ravindranath N, Venkataiah B, Ramesh C, Jayaprakash P, Das B. Jatrophene, a novel macrocyclic bioactive diterpene from *Jatropha gossypifolia*. *Chem Pharm Bull (Tokyo)*. 2003;51:870-1.
6. Akinpelu DA, Obuotor EM. Antibacterial activity of *Nicotiana tabacum* leaves. *Fitoterapia*. 2000;71:199-200.
7. Ameya G, Manilal A, Merdekios B. In vitro Antibacterial Activity and Phytochemical Analysis of *Nicotiana tabacum* L. Extracted in Different Organic Solvents. *Open Microbiol J*. 2017;11:352-359.
8. Anumudu CK, Nwachukwu MI, Obasi CC, Nwachukwu IC, Henetu FC. Antimicrobial activities of extracts of tobacco leaf (*Nicotiana tabacum*) and its grounded snuff (utaba) on *Candida albicans* and *Streptococcus pyogenes*. *J Trop Dis*. 2019;7:300.
9. Marreiro RO, Bandeira MFCL, Souza TP, Almeida MC, Bendaham K, Venâncio GN, Rodrigues IC, Coelho CN, Milério PSL, Oliveira GP, Conde NCO. Evaluation of the stability and antimicrobial activity of an ethanolic extract of *Libidibia ferrea*. *Clin Cosmet Investig Dent*. 2014;6:9–13.
10. Ferreira, DS, Rosso VV, Mercadante AZ. Bioactive compounds of blackberry fruits (*Rubus spp.*) grown in Brazil. *Rev Bras Frutic., Jaboticabal – SP*. 2010;32:664-674.
11. De-Araújo AA, Soares LA, Assunção Ferreira MR, de Souza Neto MA, da Silva GR, de Araújo RF Jr, Guerra GC, de Melo MC. Quantification of polyphenols and evaluation of antimicrobial, analgesic and anti-inflammatory activities of aqueous and acetone-water extracts of *Libidibia ferrea*, *Parapiptadenia rigida* and *Psidium guajava*. *J Ethnopharmacol*. 2014;156:88-96.

12. Akinpelu DA. Antimicrobial activity of *Anacardium occidentale* bark. *Fitoterapia*. 2001;72:286-7.
13. Baptista A, Goncalves RV, Bressan J, Peluzio M. Antioxidant and antimicrobial activities of crude extracts and fractions of cashew (*Anacardium occidentale* L.), cajui (*Anacardium microcarpum*) and pequi (*Caryocar brasiliense* c.): A systematic review. *Oxid Med Cell Longev*. 2018;2018:3753562.
14. Da-Silva CFG, Lucas AM, Santo ATE, Almeida RN, Cassel E, Vargas RMF. Sequential processing of *Psidium guajava* L. leaves: steam distillation and supercritical fluid extraction. *Braz J Chem Eng*. 2019;36:487-496.
15. Garcia S, Araiza M, Gomez M, Heredia N. Inhibition of growth, enterotoxin production, and spore formation of *Clostridium perfringens* by extracts of medicinal plants. *J Food Prot*. 2002;65:1667-9.
16. Almeida KC, Barbosa TR, Silva RNR, Jacques DS, Freire RB. Cytotoxic effect of *Psidium guajava* L. (Myrtaceae). *Rev Bras Farm*. 2006;87:60-62.
17. Sriwilajaroen N, Fukumoto S, Kumagai K, Hiramatsu H, Odagiri T, Tashiro M, Suzuki Y. Antiviral effects of *Psidium guajava* Linn. (guava) tea on the growth of clinical isolated H1N1 viruses: its role in viral hemagglutination and neuraminidase inhibition. *Antiviral Res*. 2012;94:139-46.
18. Fukumoto S, Goto T, Hayashi S, et al. Anti-sars coronavirus agent and product containing anti-sars coronavirus agent WO2010041703A1.
19. Kumar V, Jung YS, Liang PH. Anti-SARS coronavirus agents: a patent review (2008 - present). *Expert Opin Ther Pat*. 2013;23:1337-48.
20. Alnaqeeb M, Mansor KA, Mallah EM, Ghanim BY, Idkaidek N, Qinna NA. Critical pharmacokinetic and pharmacodynamic drug-herb interactions in rats between warfarin and pomegranate peel or guava leaves extracts. *BMC Complement Altern Med*. 2019;19:29.
21. Hsieh CL, Lin YC, Yen GC, Chen HY. Preventive effects of guava (*Psidium guajava* L.) leaves and its active compounds against α -dicarbonyl compounds-induced blood coagulation. *Food Chem*. 2006;103:528-535.
22. Azevedo CF, Quirino ZGM, Bruno RLA. Pharmacobotanical study of the aerial parts of Brazilian pepper (*Schinus terebinthifolius* Raddi). *Rev Bras PI Med*. 2015;17:26-35.
23. Gomes FS, Procópio TF, Napoleão TH, Coelho LC, Paiva PM. Antimicrobial lectin from *Schinus terebinthifolius* leaf. *J Appl Microbiol*. 2013;114:672-9.
24. Salem MZM, El-Hefny M, Ali HM, Elansary HO, Nasser RA, El-Settawy AAA, El Shanhorey N, Ashmawy NA, Salem AZM. Antibacterial activity of extracted bioactive molecules of *Schinus terebinthifolius* ripened fruits against some pathogenic bacteria. *Microb Pathog*. 2018;120:119-127.
25. Cole ER, dos-Santos RB, Lacerda-Júnior V, Martins JDL, Greco SJ, A. Cunha-Neto A. Chemical composition of essential oil from ripe fruit of *Schinus terebinthifolius* Raddi and evaluation of its activity against wild strains of hospital origin. *Braz J Microbiol*. 2014;45:821-828.
26. Da Costa MP, Bozinis MC, Andrade WM, Costa CR, da Silva AL, Alves de Oliveira CM, Kato L, Fernandes Ode F, Souza LK, Silva Mdo R. Antifungal and cytotoxicity activities of the fresh xylem sap of *Hymenaea courbaril* L. and its major constituent fisetin. *BMC Complement Altern Med*. 2014;14:245.
27. Golovinskaia O, Wang C. Review of functional and pharmacological activities of berries. *Molecules*. 2021;26:3904

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